Tolerance, bioavailability, and potential cognitive health implications of a distinct aqueous spearmint extract

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ABSTRACT

Background: Cognitive function can decline during the aging process and significantly reduce quality of life. Although a number of interventions have been investigated for cognitive dysfunction, including antioxidants, this prominent health concern emphasizes a need to explore methods to support cognitive health later in the life span. An aqueous extract from a proprietary spearmint line has been developed which contains a number of antioxidant compounds, including rosmarinic acid at levels that are higher than found in commercially-bred spearmint. Therefore, this pilot trial assessed the tolerance, bioavailability, and potential cognitive health implications of a proprietary spearmint extract in men and women with self-reported memory impairment.

Methods: Subjects consumed 900 mg/day spearmint extract for 30 days. The sample population (N = 11) was 73% female and 27% male with a mean age of 58.7 ± 1.6 y. Tolerability parameters were assessed at baseline and end of treatment visits. Computerized cognitive function tests were completed and blood was drawn at pre- and post-dose (0.5 to 4 h) timepoints during baseline and end of treatment visits. Subjective cognition was also assessed at end of treatment.

Results: No serious adverse events or clinically relevant findings were observed in any tolerability parameters. Plasma vanillic, caffeic, and ferulic acid sulfates, rosmarinic acid, and methyl rosmarinic acid glucuronide were detected in plasma following acute administration of the spearmint extract. Computerized cognitive function scores improved in reasoning (P =
0.023) and attention/concentration (P = 0.002) after 30 days of supplementation. After acute administration, subjects had improved attention/concentration in two tests at 2 (P = 0.042 and P = 0.025) and 4 h (P = 0.001 and P = 0.002).

Conclusions: The results from this pilot trial suggest that the spearmint extract, which contains higher rosmarinic acid content relative to extracts from typical commercial lines, was well-tolerated at 900 mg/day. In addition, the extract was bioavailable and further investigation is warranted regarding its potential for supporting cognitive health.

BACKGROUND

Cognitive function may decline in healthy individuals between early and late adulthood as a part of the aging process. The prevalence of subjective memory impairment in older patients in community-based populations is reported with a large range of variation (11-50%) [1, 2]. In a cross-sectional study of 21,024 adults 50 years of age or older, 20% of participants self-reported memory impairment [3]. An additional cross-sectional study evaluating seventeen general practice clinics that included 2,934 patients aged 65 and older reported that 23% of elderly patients surveyed self-reported memory impairment upon prompting; however, only 18% of those elderly patients who self-reported memory impairment consulted a physician for their impairment [4]. Although cognitive decline is generally accepted as a typical consequence of aging, it significantly decreases quality of life [5]. It is estimated that 5.4 million elderly Americans have cognitive impairment without dementia and roughly 12% of these individuals will develop dementia annually [6]. Although a number of treatments are available for more advanced stages of cognitive dysfunction, including dementia, there is a need to explore novel methods to prevent or slow age-associated decline in cognitive function.

Traditional medicine has long used plant-based remedies to treat a number of ailments and, more recently, plant-based dietary interventions such as ginkgo biloba, ginseng, and guarana have been investigated in clinical trials for their potential in enhancing cognitive function in healthy volunteers [7-10]. Results from several trials have suggested that consumption of plant extracts within the Lamiaceae family may promote cognitive function in healthy volunteers [11-13]. However, randomized controlled trials specifically investigating the effects of aqueous spearmint (Mentha spicata L.) extracts, a member of the Lamiaceae family, on cognitive function are limited. A few studies have been conducted evaluating the effects of small quantities of spearmint oil in spearmint chewing gum on memory in healthy volunteers, which report conflicting results [14-16]. Previous work in rodents with an aqueous spearmint extract found that 320 and 640 mg/kg body weight of the extract (16 and 32 mg rosmarinic acid/kg body weight), equivalent to 600-1200 mg of the spearmint extract on average for humans, were effective in improving memory and learning in a SAMP8 mouse model of accelerated aging [17, 18].

There are several published studies which have investigated the tolerance of consuming aqueous plant extracts within the Lamiaceae family; however a majority of these have investigated peppermint (Mentha Piperita) extracts [19]. Preclinical safety studies were conducted on the spearmint extract used in this trial in accordance with the Organisation for
Economic Cooperation and Development and the U.S. Food and Drug Administration’s RedBook 2000 guidelines [20, 21]. Genotoxicity testing results demonstrated that the spearmint extract was non-mutagenic at concentrations up to 5000 µg/plate as assessed by the Ames bacterial reverse mutation assay. In addition, the extract was non-clastogenic at dose levels up to 5000 µg/ml using the chromosomal aberration assay [22]. A 90-day toxicity study in rodents determined the No Observed Adverse Effect Level (NOAEL) to be 1948 mg/kg body weight/day of the spearmint extract, which corresponds to 300 mg rosmarinic acid/kg body weight/day [22].

Spearmint extracts are widely used as flavorings and seasonings in beverages and confectioneries and have ‘Generally Recognized as Safe’ status as a natural seasoning/flavoring, essential oil, and natural extract in the United States [23, 24]. However, spearmint extract would be consumed at doses higher than what would typically be consumed as a flavoring or seasoning, when consumed for cognitive benefit. Thus, the objective of this pilot study was to evaluate the tolerability and bioavailability of a proprietary aqueous spearmint extract when consumed at 900 mg, and its potential effects on cognitive function in healthy men and women with self-reported memory impairment.

METHODS
Chemicals
All chemicals and solvents were of analytical grade. Rosmarinic acid was purchased from Sigma-Aldrich Co. (St. Louis, MO, USA), whereas ferulic acid-4-O-sulfate and dihydrocaffeic acid 3-O-sulfate were purchased from TRC - Toronto Research Chemicals (Toronto, Ontario, Canada). All solvents and reagents were purchased from Carlo Erba Reagents (Milano, Italy). Ultrapure water from MilliQsystem (Millipore, Bedford, MA, USA) was used throughout the experiments.

Study Design
This open-label, pilot study was conducted in accordance with Good Clinical Practice Guidelines, the Declaration of Helsinki [25], and the United States 21 Code of Federal Regulations [26]. An appropriately constituted Institutional Review Board (Hummingbird IRB, Cambridge, MA) approved the study protocol and informed consent documents prior to initiation of the study [25]. This study included one telephone screen, one screening visit, and two treatment visits at the beginning and end of a 30-day treatment period. Signed informed consent and authorization for disclosure of protected health information was provided by the subjects prior to implementing any protocol-specific procedures. Subjects were informed of their right to withdraw from the study at any time.

Subjects
Generally healthy men and women 50-70 years of age, with body mass index between 18.5-35.0 kg/m², and possessing a high school diploma were recruited for this trial. Subjects were eligible for the trial if they reported gradual memory impairment by scoring ≥25 on the Memory Assessment Clinic Scale Questionnaire (MAC-Q), administered during the telephone screen [27, 28]. Eligible subjects also needed to exhibit normal cognitive function during the screening visit, as measured by a score of ≥24 on the Mini Mental State Examination (MMSE) [28, 29].
In addition, eligible subjects were those who were willing to maintain their habitual diet (including caffeinated and alcoholic beverages) and exercise routines throughout the study period, and refrain from using tobacco products (1h), consuming caffeine (10-14 h), consuming alcohol (24 h), and participating in vigorous exercise (24 h) prior to and during the test visits, as indicated by the time in parentheses.

Subjects were deemed ineligible for participation in the study based on the following exclusion criteria: uncontrolled hypertension (systolic blood pressure ≥160 mm Hg or diastolic blood pressure ≥100 mm Hg); abnormal laboratory test results of clinical significance (at the discretion of the Investigator); a history or presence of clinically important cardiac (including coronary heart disease), renal, hepatic, endocrine (including type 1 or type 2 diabetes mellitus), pulmonary, biliary, gastrointestinal, pancreatic, or neurologic disorders (including sleep disorders, head injuries; Alzheimer’s disease, Parkinson’s disease, stroke, inflammatory brain disease); a recent history or presence of cancer, except non-melanoma skin cancer; history within 12 months of screening or strong potential for alcohol (>14 servings/week) or substance abuse; history of depression within 24 months of screening; history of heavy smoking (>1 pack/day) within 3 months of screening; history of heavy caffeinated beverage consumption (>400 mg caffeine/day) within 2 weeks of screening; and history of use of psychotropic medications within one month of screening. Females who were pregnant, lactating, or planning to be pregnant during the study period or of childbearing potential and unwilling to commit to the use of a medically approved form of contraception were not enrolled. Individuals who reported occupations that resulted in disruption of sleep-wake cycles were excluded from the study. If a deviation (± 2 h) in the participants’ normal sleep duration occurred the evening before a test visit, the visit was rescheduled. In addition, individuals were excluded if use of medications or supplements known to alter cognitive function were reported within two weeks of screening. If the participant used antibiotics within 5 days of any visit or an infection occurred during the study, clinic visits were rescheduled to allow at least five days for resolution of the infection or completion of the antibiotic therapy. Finally, subjects were excluded from the study if they were unable to complete or understand the cognitive function practice tests during screening.

Study Product and Treatment

The proprietary aqueous spearmint extract (Neumentix™ Phenolic Complex K110-42) [30] was manufactured by Kemin Foods, L.C. (Des Moines, IA) and packaged in 450 mg capsules by Five-Star Pharmacy (Clive, IA). The spearmint extract contained 15.2% rosmarinic acid in addition to a number of other classes of phenolic compounds including salvianolic, caffeoylquinic, and hydroxyphenylpropanoic acids (Mena, Del Rio, et al. manuscript in preparation). Subjects were instructed to consume 900 mg (2 capsules) of the extract daily with breakfast. Compliance with study product consumption was evaluated by the study staff according to the returned quantity of study product and a study product diary that subjects completed daily. Compliance was calculated as a percentage of study product consumed.

Tolerability Assessments

Fasting (10-14 h) blood samples were collected at screening and the end of the treatment period prior to the acute administration test for plasma chemistry, whole blood hematology, and plasma
lipid profiles. Blood assessments were completed by Elmhurst Memorial Reference Laboratory (Elmhurst, IL). Plasma glucose concentrations were assessed using the glucose oxidase method. The plasma lipid profile assessment included total cholesterol (TC), high-density lipoprotein cholesterol (HDL-C), and triglycerides (TG) and was analyzed using the Beckman Coulter’s LX20 PRO (Fullerton, CA, USA) and as previously described by the Standardization Program of the Centers for Disease Control and Prevention and the National Heart, Lung, and Blood Institute [31]. The Friedewald equation was used to estimate LDL-C concentrations in mg/dL as follows: 

$$\text{LDL-C} = \text{TC} - \text{HDL-C} - \text{TG}/5.$$ 

Non-HDL-C was calculated as follows:

$$\text{non-HDL-C} = \text{TC} - \text{HDL-C}.$$ 

Vital signs (seated, resting blood pressure/heart rate) were measured twice and averaged, using an automated device, at baseline and at the end of the treatment period. Adverse events were also assessed at baseline and at the end of the treatment period, both at the beginning and end of each test visit.

A gastrointestinal (GI) tolerability questionnaire was administered at each test visit prior to test product consumption, as described previously with minor modifications [33]. Subjects were instructed to recall their GI symptoms over the last 30 d and rank them as follows: less than usual (-2), somewhat less than usual (-1), usual/not experienced (0), somewhat more than usual (1), and much more than usual (2). A composite score was also calculated for all ratings ≥0 (somewhat more than usual)

**Plasma Extraction and uHPLC-MSn Analysis**

Plasma samples were extracted as previously reported [34]. Two 0.5 mL aliquots of plasma were extracted with 1.25 mL of acetonitrile, after vortexing for 5 min. Samples were centrifuged for 10 min at 14000 rpm and 1.5 mL of supernatant was dried under vacuum, by rotary evaporation. The pellet was dissolved in 100 μL of 80% methanol in 0.1% formic acid-acidified water (v/v) and stored at -80 °C until analysis by ultra-high performance liquid chromatography coupled with tandem mass spectrometry (uHPLC-MSn).

An Accela™ uHPLC 1250 apparatus equipped with the LTQ XL™ Linear Ion Trap Mass Spectrometer (LITMS) fitted with a heated-ESI (H-ESI-II) probe (Thermo Fisher Scientific Inc., San Jose, CA, USA) was utilized for the analysis. Separations were carried out by means of a Kinetex PFP (50 x 2.1 mm), 2.6 mm particle size (Phenomenex, Torrance, CA, USA). Phenolic metabolites were detected in negative ionization mode, with mobile phase, pumped at a flow-rate of 0.3 mL/min, consisting of a mixture of acidified acetonitrile (0.1% formic acid; solvent A) and 0.1% aqueous formic acid (solvent B). Following 1 min of 2% solvent A in B, the proportion of A was increased linearly to 35% over a period of 10 min. The H-ESI-II interface was set to a capillary temperature of 275 °C and the source heater temperature was 50 °C. The sheath gas (N₂) flow rate was set at 40 (arbitrary units) and the auxiliary gas (N₂) flow rate at 5. During analysis, the source voltage was 4 kV, and the capillary voltage and tube lens voltage were -26.0 V and -77.7 V, respectively.

Initially, preliminary analysis of 5 μL of plasma extract was carried out using full-scan, data-dependent MS³, scanning from a mass to charge (m/z) of 100 to 800 using a collision-induced dissociation (CID) equal to 35 (arbitrary units) to obtain fragmentation. After this first step, further specific selected reaction monitoring (SRM) analyses were carried out to unambiguously identify and quantify the detected metabolites, by monitoring specific m/z transitions: 163→119
couchic acid), 179→135 (caffeic acid), 193→134, 149, 178 (ferulic acid), 247→147 (vanillic acid sulfate), 259→179 (caffeic acid sulfate), 261→181 (dihydrocaffeic acid sulfate), 273→193 (ferulic acid sulfate), 275→195 (dihydroferulic acid sulfate), 359→161, 179, 197 (rosmarinic acid), 369→193 (ferulic acid glucuronide), 439→359 (rosmarinic acid sulfate), 535→359 (rosmarinic acid glucuronide), 549→373 (methyl rosmarinic acid glucuronide). Molecules were fragmented using pure helium (99.99%), with a CID setting of 25 to obtain the fragmentation of the molecular ion. Identified metabolites were quantified as ferulic acid-4-\(O\)-sulfate, dihydrocaffeic acid 3-\(O\)-sulfate and rosmarinic acid equivalents by using calibration curves ranging from 1 to 1000 nmol/L. Specifically, vanillic acid sulfate, caffeic acid sulfate, ferulic acid sulfate and dihydroferulic acid sulfate were quantified as ferulic acid-4-\(O\)-sulfate equivalents, and dihydrocaffeic acid sulfate was expressed as dihydrocaffeic acid 3-\(O\)-sulfate equivalents. Rosmarinic acid and methyl rosmarinic acid glucuronide were quantified as RA equivalents.

**Cognitive Function Assessments**

Performance on a battery of publicly available computerized brain training tasks (http://www.cambridgebrainsciences.com) was utilized to assess cognitive function (Cambridge Brain Sciences, London, Ontario, Canada). These tasks were designed using established cognitive neuroscience paradigms in parallel forms to assess memory, reasoning, attention/concentration, and planning, as previously described [35]. The battery included eight tasks which are summarized in Table 1: digit span (memory 1), paired associates (memory 2), double trouble (reasoning 1), odd one out (reasoning 2), rotations (attention/concentration 1), polygons (attention/concentration 2), spatial search (planning 1), and spatial slider (planning 2). Subjects completed a practice session during the screening visit to gain familiarity with the tests, to limit training effects and variability in the test scores, and to ensure an optimal level of performance. The practice sessions included two complete test batteries separated by at least one hour. The battery was administered at -1, 2.25, and 4 h, where \( t = 0 \) h was the study product/breakfast consumption at baseline and end of treatment, respectively. Each battery took approximately 30-45 min and was completed in an environment where temperature, lighting, and noise were kept constant.

The Subject Global Impression (SGI) Scale of Cognition questionnaire was administered at the end of the 30-day supplementation period to assess subjective cognition [28]. Subjects were instructed to rate their memory, attention, and speed of thinking at the end of the study relative to their condition prior to inclusion in the study as follows: very much improved (1), much improved (2), minimally improved (3), no change (4), minimally worse (5), much worse (6), and very much worse (7). A composite score was calculated as the average of the three domains (memory, attention, and speed of thinking). The difference from ‘no change’ (4) was calculated for each domain and the composite score.

**Table 1.** Overview of the cognitive function test battery
d

<table>
<thead>
<tr>
<th>Task</th>
<th>Domain/Designation</th>
<th>Brief Description</th>
<th>Outcome Measure</th>
</tr>
</thead>
</table>


The cognitive function test battery included eight publicly available (http://www.cambridgebrainsciences.com) tasks (Cambridge Brain Sciences, London, Ontario, Canada).

Statistical Analysis
A sample of 11 subjects was enrolled and no formal sample size calculations were completed since this study was a pilot trial. Statistical analyses were completed using SAS version 9.2 (SAS Institute, Cary, NC). All tests of significance in this pilot study were performed at alpha <0.1, two-sided in this exploratory pilot trial. P-values were calculated using a paired t-test if the data were normally distributed or the Wilcoxon sign-rank test if the normality assumption was rejected at the 1% level with the Shapiro-Wilk test [36]. Statistical analyses for safety measures were completed for data collected from all subjects who were randomized and consumed at least one dose of study product. The analyses of outcomes were completed on a modified intention-to-treat (MITT) sample. The MITT sample included all subjects who provided at least one post-randomization outcome data point during the treatment. In addition, a per protocol (PP) sample comprised a subset of the MITT population. Subjects were excluded from the PP sample for violations of inclusion/exclusion criteria and non-compliance including: missing appointments, <80% or >120% compliance with study product consumption, or failure to consume the entire
study product at any test visit. Data are reported as means ± standard error of the mean (SEM) or median with interquartile limits for the MITT sample. Differences between the MITT and PP samples are noted if present.

The co-primary outcome variables were the changes from baseline to the end of treatment for the GI tolerability questionnaire composite score and the SGI questionnaire overall composite score, both of which reflected chronic dosing effects. Secondary outcome variables included differences between baseline and the end of treatment for individual scores of the GI tolerability questionnaire (nausea, gas/bloating, flatulence, cramping, and diarrhea/loose stools), the SGI questionnaire (memory, attention, speed of thinking) and the computerized cognitive function tasks (two tasks for each domain; attention/concentration, memory, planning, reasoning; Table 1). Cognitive function scores were evaluated for acute (-1 to 2.25 and 4 h at baseline) and chronic (baseline to end of treatment using the -1 h assessment) differences. Exploratory outcome variables were evaluated both acutely at the baseline test visit (day 0; differences between -1.25 and both 0.5 and 2 h assessments) and chronically (differences between baseline and end of treatment using the -1.25 h assessment) in free and conjugated plasma rosmarinic acid metabolites.

RESULTS

Subjects
In total, 20 participants were screened for this trial and 11 eligible subjects were identified. Of the 11 subjects who were enrolled in the study, one subject withdrew consent after the baseline test visit due to an inability to understand the cognitive function tests and was removed from the PP sample. A second subject was removed from the PP sample due to 134% compliance for study product consumption. Therefore, the MITT sample included 11 subjects at baseline and 10 completers, and the PP sample included 9 individuals at each assessment. A single adverse event, back pain, was reported during the treatment period and coded as unrelated to the study product consumption.

Baseline characteristics of the MITT sample (N = 11) are included in Table 2.

Table 2. Baseline characteristics of subjects in the modified intention-to-treat sample

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Overall Value (N = 11)</th>
</tr>
</thead>
<tbody>
<tr>
<td>n (%)</td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>3 (27)</td>
</tr>
<tr>
<td>Female</td>
<td>8 (73)</td>
</tr>
<tr>
<td>Race/Ethnicity</td>
<td></td>
</tr>
<tr>
<td>Non-Hispanic White</td>
<td>10 (91)</td>
</tr>
<tr>
<td>Black/African American</td>
<td>1 (9)</td>
</tr>
<tr>
<td>Mean (SEM)</td>
<td></td>
</tr>
<tr>
<td>Age (years)</td>
<td>58.7 (1.6)</td>
</tr>
<tr>
<td>Body mass index (kg/m²)</td>
<td>27.4 (1.0)</td>
</tr>
<tr>
<td>MAC-Q score</td>
<td>29.7 (1.0)</td>
</tr>
<tr>
<td>MMSE score</td>
<td>28.9 (0.4)</td>
</tr>
</tbody>
</table>

Abbreviations: MAC-Q, Memory Assessment Clinic Scale Questionnaire; MMSE, Mini-Mental State Exam
The sample was comprised of 27% males and 73% females, with mean age and BMI of 58.7 ± 1.6 y and 27.4 ± 1.0 kg/m², respectively. Mean overall compliance with study product consumption was 103.2 ± 3.3%. Mean scores for the qualifying MAC-Q and MMSE were 29.7 ± 1.0 and 28.9 ± 0.4, respectively.

**Tolerability**

Consumption of the spearmint extract did not significantly alter individual GI symptoms (constipation, cramping, flatulence, gas/bloating, loose stools, and nausea) between baseline and the end of treatment (P = 1.000 for all comparisons; data not shown). In addition, the GI tolerability composite score did not change significantly between baseline and the end of treatment (P = 1.000; data not shown).

Mean and median values for vital signs and fasting lipoprotein lipids, at both baseline and end of treatment and the change from baseline, are presented in Table 3.

**Table 3.** Vital signs and fasting lipoprotein lipids at baseline, end of treatment, and change from baseline in response to spearmint supplementation

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Baseline¹</th>
<th>EOT²</th>
<th>Difference (Δ)</th>
<th>P-value³</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean (SEM) or Median (Interquartile Limits)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>SBP (mm Hg)</td>
<td>121.1 (3.6)</td>
<td>121.7 (3.3)</td>
<td>-0.9 (2.4)</td>
<td>0.706</td>
</tr>
<tr>
<td>DBP (mm Hg)</td>
<td>75.3 (2.5)</td>
<td>78.3 (2.6)</td>
<td>1.3 (2.3)</td>
<td>0.603</td>
</tr>
<tr>
<td>Heart rate (bpm)</td>
<td>63.2 (2.2)</td>
<td>68.0 (2.7)</td>
<td>3.7 (1.8)</td>
<td><strong>0.077</strong></td>
</tr>
<tr>
<td>Body weight (kg)</td>
<td>77.1 (2.6)</td>
<td>77.8 (2.9)</td>
<td>0.4 (0.3)</td>
<td>0.212</td>
</tr>
<tr>
<td>LDL-C (mg/dL)</td>
<td>138.6 (11.5)</td>
<td>148.6 (11.4)</td>
<td>5.2 (5.4)</td>
<td>0.361</td>
</tr>
<tr>
<td>Non-HDL-C (mg/dL)</td>
<td>156.6 (10.8)</td>
<td>163.7 (11.6)</td>
<td>3.1 (5.5)</td>
<td>0.584</td>
</tr>
<tr>
<td>TC (mg/dL)</td>
<td>213.1 (11.5)</td>
<td>222.3 (12.5)</td>
<td>3.4 (6.0)</td>
<td>0.586</td>
</tr>
<tr>
<td>HDL-C (mg/dL)</td>
<td>56.6 (3.4)</td>
<td>58.6 (3.2)</td>
<td>0.3 (1.6)</td>
<td>0.858</td>
</tr>
<tr>
<td>Triglycerides (mg/dL)</td>
<td>88.2 (8.8)</td>
<td>75.7 (6.1)</td>
<td>-9.3 (8.3)</td>
<td>0.293</td>
</tr>
<tr>
<td>TC/HDL-C</td>
<td>3.7 (3.1, 4.3)</td>
<td>3.6 (3.4, 4.5)</td>
<td>0.1 (-0.1, 0.3)</td>
<td>0.432</td>
</tr>
</tbody>
</table>

Abbreviations: bpm, beats per minute; DBP, diastolic blood pressure; HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol; SBP, systolic blood pressure; TC, total cholesterol.

¹Baseline refers to pre-dose values on day 0 (N = 11).
²End of treatment (EOT) refers to pre-dose values on day 30 (n = 10).
³P-values were calculated from paired t-tests or Wilcoxon sign rank test, between baseline and end of treatment in the modified intention-to-treat sample.

No significant differences in lipid parameters were evident over the 30 day treatment period in the MITT sample. An increase in LDL cholesterol in the PP sample was evident over the treatment period (137.0 ± 10.7 vs. 145.89 ± 12.4 mg/dL; P = 0.079). Heart rate increased slightly over the 30 day treatment period (63.2 ± 2.2 vs. 68.0 ± 2.6 bpm; P = 0.077) in the MITT population; however, this change was no longer significant in the PP sample (P = 0.155). Body
weight increased over the 30 day treatment period in the PP sample by \((77.0 \pm 3.2 \text{ vs. } 77.6 \pm 3.3 \text{ kg; } P = 0.062)\), but not in the MITT sample. Blood chemistry and hematology values at baseline and end of treatment, and the change from baseline are presented in Tables 4 and 5.

### Table 4. Plasma chemistry panel values at baseline, end of treatment, and changes from baseline, in response to spearmint supplementation

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Baseline(^1)</th>
<th>EOT(^2)</th>
<th>Difference ((\Delta))</th>
<th>P-value(^3)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mean (SEM)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Glucose (mg/dL)</td>
<td>94.9 (1.5)</td>
<td>97.3 (2.1)</td>
<td>1.6 (1.5)</td>
<td>0.318</td>
</tr>
<tr>
<td>Sodium (mmol/L)</td>
<td>140.3 (0.4)</td>
<td>140.2 (0.3)</td>
<td>-0.1 (0.5)</td>
<td>0.847</td>
</tr>
<tr>
<td>Potassium (mmol/L)</td>
<td>4.4 (0.1)</td>
<td>4.6 (0.1)</td>
<td>0.1 (0.2)</td>
<td>0.447</td>
</tr>
<tr>
<td>Chloride (mmol/L)</td>
<td>104.9 (0.6)</td>
<td>105.5 (0.8)</td>
<td>0.2 (0.9)</td>
<td>0.836</td>
</tr>
<tr>
<td>Carbon dioxide (mmol/L)</td>
<td>29.5 (0.4)</td>
<td>29.8 (0.6)</td>
<td>0.6 (0.4)</td>
<td>0.193</td>
</tr>
<tr>
<td>BUN (mg/dL)</td>
<td>13.6 (0.8)</td>
<td>14.1 (1.0)</td>
<td>0.4 (0.8)</td>
<td>0.637</td>
</tr>
<tr>
<td>Creatinine (mg/dL)</td>
<td>0.9 (0.0)</td>
<td>0.9 (0.1)</td>
<td>0.0 (0.0)</td>
<td>0.515</td>
</tr>
<tr>
<td>BUN/Creatinine</td>
<td>15.7 (1.2)</td>
<td>16.4 (1.8)</td>
<td>0.5 (1.6)</td>
<td>0.785</td>
</tr>
<tr>
<td>Anion gap (mmol/L)</td>
<td>5.9 (0.5)</td>
<td>4.9 (0.4)</td>
<td>-0.9 (0.4)</td>
<td>0.068</td>
</tr>
<tr>
<td>Calcium (mg/dL)</td>
<td>9.6 (0.1)</td>
<td>9.4 (0.1)</td>
<td>-0.2 (0.0)</td>
<td>0.007</td>
</tr>
<tr>
<td>Calcium osmolality (mOs/kg)</td>
<td>290.6 (0.8)</td>
<td>290.7 (0.8)</td>
<td>-0.1 (1.1)</td>
<td>0.931</td>
</tr>
<tr>
<td>AST (U/L)</td>
<td>22.2 (1.5)</td>
<td>22.6 (1.8)</td>
<td>0.4 (1.4)</td>
<td>0.786</td>
</tr>
<tr>
<td>ALT (U/L)</td>
<td>20.3 (1.7)</td>
<td>19.8 (2.0)</td>
<td>-0.6 (2.1)</td>
<td>0.776</td>
</tr>
<tr>
<td>ALP (U/L)</td>
<td>69.9 (5.7)</td>
<td>66.7 (5.6)</td>
<td>-1.5 (1.0)</td>
<td>0.169</td>
</tr>
<tr>
<td>Total bilirubin (mg/dL)</td>
<td>0.8 (0.1)</td>
<td>0.7 (0.1)</td>
<td>-0.0 (0.1)</td>
<td>0.790</td>
</tr>
<tr>
<td>Total protein (g/dL)</td>
<td>6.9 (0.2)</td>
<td>6.7 (0.2)</td>
<td>-0.2 (0.1)</td>
<td>0.055</td>
</tr>
<tr>
<td>Albumin (g/dL)</td>
<td>4.0 (0.1)</td>
<td>3.9 (0.1)</td>
<td>-0.1 (0.1)</td>
<td>0.111</td>
</tr>
<tr>
<td>Globulin (g/dL)</td>
<td>2.9 (0.2)</td>
<td>2.9 (0.2)</td>
<td>-0.0 (0.1)</td>
<td>0.421</td>
</tr>
</tbody>
</table>

Abbreviations: ALP, alkaline phosphatase; ALT, alanine transaminase; AST, aspartate aminotransferase; BUN, blood urea nitrogen; EOT, end of treatment.

1\(^{Baseline}\) refers to pre-dose values on day 0 (N = 11).

2\(^{End of treatment}\) refers to pre-dose values on day 30 (n = 10).

3\(^{P-values\) were calculated from paired t-tests or Wilcoxon sign rank test, between baseline and end of treatment in the modified intention-to-treat sample.

Values from the blood chemistry panel in the MITT sample revealed declines in calcium (9.6 ± 0.1 vs. 9.4 ± 0.1 mg/dL, P = 0.007), the anion gap (5.9 ± 0.5 vs. 4.9 ± 0.4 mmol/L, P = 0.068), and total protein (6.9 ± 0.2 vs. 6.7 ± 0.2 mg/dL, P = 0.055) between baseline and end of treatment, respectively. These differences were also evident in the PP sample. However, the noted differences in heart rate, LDL cholesterol, body weight, and blood chemistry values are within normal ranges and biological variability, thus not considered clinically relevant. In
addition, no significant differences were evident over the 30 day treatment for whole blood hematology values.

**Table 5.** Whole-blood hematology panel values at baseline, end of treatment, and changes from baseline, in response to spearmint supplementation

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Baseline&lt;sup&gt;1&lt;/sup&gt;</th>
<th>EOT&lt;sup&gt;2&lt;/sup&gt;</th>
<th>Difference (Δ)</th>
<th>P-value&lt;sup&gt;3&lt;/sup&gt;</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean (SEM) or Median (Interquartile limits)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>WBC (cells/μL)</td>
<td>5.6 (4.1, 6.9)</td>
<td>4.6 (4.2, 6.3)</td>
<td>0.3 (-0.1, 0.4)</td>
<td>0.447</td>
</tr>
<tr>
<td>RBC (cells x 10&lt;sup&gt;6&lt;/sup&gt;/μL)</td>
<td>4.5 (0.1)</td>
<td>4.6 (0.1)</td>
<td>0.1 (0.1)</td>
<td>0.529</td>
</tr>
<tr>
<td>Hemoglobin (g/dL)</td>
<td>13.4 (0.2)</td>
<td>13.6 (0.3)</td>
<td>0.2 (0.3)</td>
<td>0.529</td>
</tr>
<tr>
<td>Hematocrit (%)</td>
<td>39.9 (0.7)</td>
<td>40.4 (0.9)</td>
<td>0.4 (0.9)</td>
<td>0.634</td>
</tr>
<tr>
<td>MCV (fL)</td>
<td>88.4 (1.6)</td>
<td>88.5 (1.7)</td>
<td>-0.0 (0.4)</td>
<td>0.980</td>
</tr>
<tr>
<td>MCH (pg/cell)</td>
<td>29.7 (0.7)</td>
<td>29.8 (0.7)</td>
<td>0.1 (0.2)</td>
<td>0.648</td>
</tr>
<tr>
<td>Platelets (cells x 10&lt;sup&gt;3&lt;/sup&gt;/μL)</td>
<td>221.7 (8.6)</td>
<td>222.5 (11.2)</td>
<td>0.3 (2.5)</td>
<td>0.906</td>
</tr>
<tr>
<td>Neutrophils (cells x 10&lt;sup&gt;3&lt;/sup&gt;/μL)</td>
<td>3.1 (0.3)</td>
<td>3.0 (0.3)</td>
<td>-0.0 (0.2)</td>
<td>0.916</td>
</tr>
<tr>
<td>Lymphocytes (cells x 10&lt;sup&gt;3&lt;/sup&gt;/μL)</td>
<td>1.7 (0.1)</td>
<td>1.7 (0.2)</td>
<td>0.0 (0.1)</td>
<td>0.713</td>
</tr>
<tr>
<td>Monocytes (cells x 10&lt;sup&gt;3&lt;/sup&gt;/μL)</td>
<td>0.5 (0.0)</td>
<td>0.5 (0.1)</td>
<td>0.0 (0.0)</td>
<td>0.193</td>
</tr>
<tr>
<td>Eosinophils (cells x 10&lt;sup&gt;3&lt;/sup&gt;/μL)</td>
<td>0.1 (0.1, 0.2)</td>
<td>0.1 (0.1, 0.2)</td>
<td>0.0 (0.0, 0.0)</td>
<td>0.500</td>
</tr>
<tr>
<td>Basophils (cells x 10&lt;sup&gt;3&lt;/sup&gt;/μL)</td>
<td>0.1 (0.0, 0.1)</td>
<td>0.1 (0.0, 0.1)</td>
<td>0.0 (0.0, 0.0)</td>
<td>1.000</td>
</tr>
</tbody>
</table>

Abbreviations: EOT, end of treatment; MCH, mean corpuscular hemoglobin; MCV, mean corpuscular volume; RBC, red blood cells; WBC, white blood cells.

<sup>1</sup>Baseline refers to pre-dose values on day 0 (N = 11).

<sup>2</sup>End of treatment (EOT) refers to pre-dose values on day 30 (n = 10).

<sup>3</sup>P-values were calculated from paired t-tests or Wilcoxon sign rank test, between baseline and end of treatment in the modified intention-to-treat sample.

### Plasma Rosmarinic Acid Metabolites

Mean concentrations of plasma rosmarinic acid metabolites following acute administration of spearmint during the baseline test visit are shown in Figure 1. Among the SRM transition analyses, a number of metabolites were detected following acute administration including vanillic acid sulfate, caffeic acid sulfate, dihydrocaffeic acid sulfate, ferulic acid sulfate, dihydroferulic acid sulfate and rosmarinic acid. A compound with a m/z equal to 549, which fragmented producing an ion with a m/z of 373, was likely methyl rosmarinic acid glucuronide, although the standard compound was not available. Vanillic acid sulfate, caffeic acid sulfate and ferulic acid sulfate were significantly elevated in plasma at 0.5 h by 124, 168, 150%, and at 2 h by 181, 219, 260%, respectively, relative to the pre-dose assessment at baseline (P < 0.02 for all comparisons). At baseline, rosmarinic acid was undetectable in plasma drawn at the pre-dose assessment but was significantly elevated in plasma by 2 h after supplement consumption (11.5 ±
4.89 nM, \( P = 0.016 \)) but not at the 0.5 h assessment (\( P = 0.250 \)). Mean plasma methyl rosmarinic acid glucuronide was also significantly elevated at 0.5 h following supplement consumption, relative to the pre-dose assessment (30.2 ± 4.4 vs. 23.3 ± 4.9 nM, \( P = 0.034 \)). However, this difference was no longer significant at the 2 h assessment (\( P = 0.289 \)). Due to the relatively high concentrations of dihydrocaffeic acid sulfate and dihydroferulic acid sulfate at the pre-dose assessment, likely related to the consumption of their precursors in coffee and wheat-based products, no differences were detected at 0.5 or 2 h after supplementation at baseline (\( P > 0.1 \)) [37, 38].

The chronic assessment of the plasma rosmarinic acid metabolites following 30 days of spearmint supplementation are presented in Figure 2. Methyl rosmarinic acid glucuronide was elevated in the plasma over the 30 day intervention, relative to the baseline assessment (30.1 ± 4.0 vs. 23.3 ± 4.88 nM, \( P = 0.082 \)). This difference was no longer significant in the PP sample (\( P = 0.107 \)). However, further investigation is warranted due to the lack of an available standard and the unexpectedly high levels detected at baseline. Mean plasma vanillic acid sulfate was reduced over the 30 day supplementation period, relative to baseline (2.6 ± 0.6 vs. 4.2 ± 0.9 nM, \( P = 0.037 \)). However, no significant differences were evident in any of the additional

**Figure 1. Acute assessment of plasma rosmarinic acid metabolites following spearmint supplementation.** Subjects (\( n = 10 \)) consumed 900 mg/day spearmint extract and blood was drawn pre-dose (-1.25 h) and post-dose (0.5 and 2 h). Vanillic acid sulfate, caffeic acid sulfate, ferulic acid sulfate and dihydroferulic acid sulfate were quantified as ferulic acid-4-O-sulfate equivalents; dihydrocaffeic acid sulfate was expressed as dihydrocaffeic acid 3-O-sulfate equivalents. Rosmarinic acid (RA) and methyl RA glucuronide were quantified as RA equivalents. Bars represent mean scores ± SEM. \( P \)-values were calculated between pre-dose (-1.25 h) and post-dose (0.5 and 2 h) assessments using a paired t-test or the Wilcoxon signed-rank test. *Significant difference (\( P < 0.05 \)). Abbreviation: RA, rosmarinic acid.
metabolites detected (caffeic acid sulfate, dihydrocaffeic acid sulfate, ferulic acid sulfate, and rosmarinic acid) following chronic supplementation.

**Cognitive Function**

Mean scores from the acute assessment of cognitive function using the computerized brain training tasks administered during the baseline test visit are represented in Figure 3. Acute cognitive function evaluation at baseline suggests improvement in mean scores for attention/concentration 1 task from pre-dose to 2.25 and 4 h post-dose by 30% (19.0 ± 8.2 points, \( P = 0.042 \)) and 46% (29.1 ± 6.6 points, \( P = 0.001 \)), relative to the pre-dose assessment, respectively. Similarly, mean scores from the attention/concentration 2 task increased between the pre-dose and 2.25 and 4 h post-dose assessments by 93% (16.8 ± 6.4 points, \( P = 0.025 \)) and 121% (21.8 ± 5.3 points, \( P = 0.002 \)), respectively. Mean scores from the planning 2 task were
also significantly elevated by 39% (11.7 ± 3.2 points; P = 0.004) at the 4 h post-dose, relative to the pre-dose assessment.

The chronic assessment of cognitive function using the computerized brain training task scores administered at the pre-dose timepoints (-1 h) during the baseline and end of treatment visits, are represented in Figure 4. Scores from reasoning 1, attention/concentration 2, and planning 2 cognitive function tasks improved between baseline and the end of treatment by 35% (6.4 ± 4.2 points; P = 0.023), 125% (22.9 ± 5.3 points; P = 0.002), and 48% (11.3 ± 5.9; P = 0.088), relative to the pre-dose assessment (t = -1 h), respectively. Scores from reasoning 1 and attention/concentration 2 remained significant (P = 0.030 and P = 0.004, respectively), the change in the planning 2 task score was no longer significant (P = 0.169) in the PP sample. All other chronic assessment scores from the cognitive function tasks did not differ significantly between baseline and end of treatment.

Mean scores from the SGI Questionnaire, which assessed change from baseline in three domains of cognition (memory, attention, and speed of thinking), are shown in Table 6. A modest improvement was evident in the average composite score from the SGI Questionnaire (3.5 ± 0.3 vs. a score of 4 representing ‘no change’; P = 0.063) after 30 days of supplementation. The difference in the average composite score (from a score of 4), was no longer significant in the PP sample (P = 0.125). There were no significant differences in individual ratings from the SGI Questionnaire.
Table 6. Subject Global Impression (SGI) scale of cognition questionnaire scores at the end of treatment in response to spearmint supplementation

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Mean (SEM)</th>
<th>P-value$^1$</th>
</tr>
</thead>
<tbody>
<tr>
<td>Memory</td>
<td>3.7 (0.2)</td>
<td>0.500</td>
</tr>
<tr>
<td>Attention</td>
<td>3.5 (0.2)</td>
<td>0.125</td>
</tr>
<tr>
<td>Speed of thinking</td>
<td>3.4 (0.3)</td>
<td>0.125</td>
</tr>
<tr>
<td>Average score</td>
<td>3.5 (0.2)</td>
<td><strong>0.063</strong></td>
</tr>
</tbody>
</table>

Abbreviations: SEM, standard error of the mean.

$^1$The SGI questionnaire was administered at the end of the 30 d treatment and subjects were asked to compare their current condition to their condition prior to inclusion in the study. Scores were coded as: 1 = very much improved, 2 = much improved, 3 = minimally improved, 4 = no change, 5 = minimally worse, 6 = much worse, 7 = very much worse.

$^2$P-values were calculated from Wilcoxon sign rank test, testing the difference from 4 (no change; n = 10) at the end of treatment in the modified intention-to-treat sample.

**DISCUSSION**

A limited number of previously published studies have evaluated the tolerance of aqueous spearmint extracts in humans at dose levels that exceed what would typically be consumed as a seasoning or flavoring. Female subjects with hirsutism (N = 21; mean age = 22 y) were enrolled in a study to evaluate the antiandrogenic effects of spearmint tea [39]. Subjects consumed 500 mg/day spearmint extract and completed a battery of publicly available cognitive function tasks (Cambridge Brain Sciences, London, Ontario, Canada; http://www.cambridgebrainsciences.com) to chronically assess memory, reasoning, attention/concentration, and planning at baseline (day 0) and the end of treatment (day 30) at the pre-dose assessment (t = -1 h). The battery included eight tasks: digit span (memory 1), paired associates (memory 2), double trouble (reasoning 1), odd one out (reasoning 2), rotations (attention/concentration 1), polygons (attention/concentration 2), spatial search (planning 1), and spatial slider (planning 2). Bars represent mean scores ± SEM. P-values were calculated between baseline and end of treatment using a paired t-test or the Wilcoxon signed-rank test.
mL of tea/day for five days, prepared with 10 g spearmint [40]. Free testosterone was significantly reduced and luteinizing hormone and follicle stimulating hormone levels increased, relative to baseline (P < 0.05), following spearmint tea consumption. In addition, biochemical tolerance parameters were assessed including plasma glucose, hepatic enzymes, and lipids. A reduction in triglycerides was the only significant (P < 0.05) finding over the brief supplementation period and was not confirmed after 30 days of supplementation in this study.

A few studies have evaluated spearmint toxicity in animal models. Specifically, in a study by Akdogan et al. [41], rats (n = 12/group) were fed spearmint tea (20 and 40 g/L) *ad libitum* or the vehicle water for 30 days. Plasma concentrations of urea and creatinine were significantly elevated (P < 0.003) at both dose levels, relative to the control, following spearmint tea consumption. Similarly, a second study utilizing the same study design in rats also reported significant elevations in activity of hepatic enzymes at both dose levels, aspartate aminotransferase (AST) and alanine aminotransferase (ALT), relative to control (P < 0.016)[42]. Spearmint intake was estimated at 2.2 g/kg body weight (20 g/L) and 4.4 g/kg body weight (40 g/L) per day in these studies, which roughly translates to a 25-50 g/day dose in a 70 kg human [18]. The estimated levels of spearmint consumption in these animal studies were ~3-fold higher per kg body weight than consumed in the current study, and no elevations were observed among participants in urea, creatinine, or hepatic enzymes.

Little is known regarding the bioavailability of aqueous spearmint extracts in humans. In a study by Baba et al. [43], rosmarinic acid was orally administered to rats at 50 mg/kg body weight. Rosmarinic acid, methyl rosmarinic acid, and coumaric acid were detected in the plasma largely as sulfated and glucuronidated conjugates. After oral administration, peak concentrations were reported at 0.5, 1, and 8 h for rosmarinic acid, methyl rosmarinic acid, and coumaric acid, respectively. In a follow-up study, the aqueous Lamiaceae extract, *Perilla frutescens*, containing 200 mg rosmarinic acid (w/w) or placebo were administered to healthy male subjects (mean age = 37 y) in a crossover design [44]. Rosmarinic, methyl rosmarinic, and ferulic acids were detected in plasma. These metabolites were present mainly as conjugates (glucuronidated and sulfated) rather than in their free form, similar to the results reported here, with peak concentrations at 0.5, 2, and 0.5 h for rosmarinic, methyl rosmarinic, and ferulic acids, respectively, following consumption of the *Perilla frutescens* extract. In contrast, rosmarinic acid sulfate, rosmarinic acid glucuronide, methyl rosmarinic acid, ferulic acid and ferulic acid glucuronide were not detected in the plasma of subjects following supplement consumption in this study, while vanillic acid sulfate, caffeic acid sulfate, dihydrocaffeic acid sulfate and dihydroferulic acid sulfate were present. No significant differences were evident in dihydrocaffeic acid sulfate and dihydroferulic acid sulfate following acute spearmint consumption in this study, which could partially be explained by the contribution of the colonic microflora to the metabolism of other phenolic compounds from other sources in the diet, such as grains and coffee [37, 38]. Interindividual differences in colonic microflora may also account for the variation in plasma concentrations of these metabolites. It is plausible that rosmarinic acid, a hydroxycinnamic acid and caffeic acid ester, may be cleaved in the small intestine and further metabolized and/or conjugated prior to absorption, similar to findings reported following consumption of other hydroxycinnamic acids (e.g. cholorgenic acids) [45, 46].
Conflicting evidence exists regarding the effects of spearmint on cognitive function and are limited to spearmint chewing gum specifically. Mint chewing gum is commonly formulated with oil extracts and contains small amounts of extract, typically 0.1-5% (w/w), but the composition and quantity of spearmint extract used in these trials were not described \[14, 15\]. Further, the interpretation of these results is difficult given the lack dosing information and it is uncertain if the improvement in memory and attention/concentration is the result of spearmint or the act of chewing, as a number of studies suggest the act of chewing alone may support cognitive function \[47\]. In the current study, acute supplementation with the spearmint extract was associated with improvements in attention/concentration tasks but did not lead to improvements in memory.

It should be noted that several other studies have been conducted which investigated the effects of acute consumption of other extracts of plants within the Lamiaceae family on cognitive function. However, a majority of these studies have evaluated the effects of either extracts of the essential oil fractions or the dried leaves of the plant. Specifically, randomized crossover trials indicate Spanish sage (Salvia lavandulaeefolia) oil acutely improves memory up to 6 h after supplementation in healthy young \(\text{age} = 18-37\) y participants, but no differences were evident in attention/concentration, compared to control \[48, 49\]. A follow-up study using a different cognitive function test battery in young adults \(\text{mean age} = 23.8\) y suggests improvements in both memory and attention/concentration up to 4 h post-supplementation following an acute dose of Spanish sage oil, relative to control. In older subjects \(\text{mean age} = 73.0\) y; \(n = 24\), both memory and attention/concentration were improved up to 6 h post-supplementation following an acute 333 mg dose of ethanol-extracted sage (Salvia officinalis) extract, relative to placebo \[12\]. Lemon balm (Melissa officinalis) improved memory at 1, 3, and 6 h following consumption of 1600 mg of dried leaf extract in healthy young adults \(\text{mean age} = 19\) y\[50\]. Similarly, dried rosemary (Rosmarinus officinalis) extract improved memory within 6 h of an acute 750 mg dose \(\text{P} = .01\) in elderly subjects \(\text{mean age} = 75\) y, but no differences were reported in attention/concentration, relative to placebo \[13\].

Compounds in plants within the Lamiaceae family are likely responsible for the wide range of reported biological activity of these plant extracts \[51\]. Rosmarinic acid is one of the phenolic compounds in plants within the Lamiaceae family that may contribute to the reported activities of these extracts, including antioxidant and anticholinesterase activities \[52\]. The spearmint extract utilized in this trial contained 15% rosmarinic acid, which significantly exceeds the typical 0-6% present in most extracts of Lamiaceae species and extracts from traditional spearmint lines grown for flavoring applications \[30, 53, 54\]. The antioxidant capacity of rosmarinic acid and/or its derived metabolites may also contribute to the observed effects of this spearmint extract on cognitive function. Acetylcholinesterase inhibitors (e.g., donepezil) and antioxidants (e.g., vitamin E) have been investigated in clinical trials for mild cognitive impairment and more advanced stages of cognitive dysfunction, including Alzheimer’s disease \[55, 56\]. Rosmarinic acid fractions of lemon balm extracts have been shown to significantly inhibit acetylcholinesterase in a time- and dose-dependent manner \[57\]. Treatment with rosmarinic acid at 10 mg/kg body weight for 21 days significantly reduced acetylcholinesterase activity in the hippocampus, cortex, and striatum of diabetic rats, relative to placebo-treated diabetic animals \[58\]. In addition to rosmarinic acid, other phenolic compounds detected in the
spearmint extract (Mena, Del Rio, et al. *manuscript in preparation*) have been reported to have antioxidant activity and could potentially be contributing to the observed cognitive benefits as well [59, 60]. Furthermore, results of a recent systematic review of observational studies show the potential for protective effects of antioxidant nutrients against age-related cognitive decline, suggesting that further investigation is warranted [61].

While the current study exhibits strengths, there are also limitations due to the lack of both a placebo control and blinding of participants to the treatment. This could have contributed to bias particularly in the self-reported outcomes. Additional limitations of the present work include that participants were free-living, consequently confounding by other dietary and/or lifestyle factors was possible. Although participants were asked to keep the number of hours they slept constant, the quality of sleep was not evaluated and may have contributed to cognitive performance [62, 63]. Training effects and intentional poor performance can be limitations to studies designed with cognitive function testing [64, 65]. Participants did complete practice tests to reduce variability in the cognitive function test scores, all tests were completed under supervision, and parallel versions were available to control for this limitation. Although without the inclusion of a control group it is difficult to rule out the effects of practice. Finally, the current trial investigated the tolerance of the spearmint extract in a small sample which included only 50-70 y old subjects with subjective memory impairment; as a result, further examination is warranted.

**CONCLUSIONS**

This open-label, pilot trial demonstrated that consumption of the aqueous spearmint extract containing at 900 mg/day for 30 days was well-tolerated and bioavailable. Although differences were evident in LDL cholesterol, anion gap, calcium, total protein, heart rate, and body weight, the observed differences were within normal biological variability, thus not considered clinically relevant. Plasma vanillic, caffeic, and ferulic acid sulfates, rosmarinic acid, and methyl rosmarinic acid glucuronide were detected in plasma within 2 h of aqueous spearmint extract administration. Moreover, methyl rosmarinic acid glucuronide was significantly elevated in plasma after 30 days of supplementation. The results of this trial suggest that chronic supplementation with the aqueous spearmint extract may impact cognitive function domains including reasoning, attention/concentration, and planning, while acute intake may have positive effects on attention/concentration and planning, over a 30-day supplementation period, relative to baseline.

In conclusion, the aqueous spearmint extract, containing higher rosmarinic acid content relative to extracts from typical commercial lines, was well-tolerated and bioavailable in older subjects (50-70 y) with self-reported memory impairment. In addition, the aqueous spearmint extract may have implications in cognitive health and warrants further investigation. Moreover, the results of this trial are essential in guiding the design of follow-up randomized controlled trials to evaluate the effects of the aqueous spearmint extract on cognitive function.

**Abbreviations:** AST, aspartate aminotransferase; ALT, alanine aminotransferase; bpm, beats per minute; BUN, blood urea nitrogen; DBP, diastolic blood pressure; EOT, end of treatment; GI, gastrointestinal; HDL-C, high-density lipoprotein cholesterol; MAC-Q, Memory Assessment
Clinic Scale Questionnaire; MCH, mean corpuscular hemoglobin; MCV, mean corpuscular volume; MMSE, Mini Mental State Examination; MITT, modified intention-to-treat; PP, per protocol; RA, rosmarinic acid; RBC, red blood cells; SBP, systolic blood pressure; SGI, Subject Global Impression; TC, total cholesterol; TG, triglycerides WBC, white blood cells.

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43. Baba S, Osakabe N, Natsume M, Terao J. Orally administered rosmarinic acid is present as the conjugated and/or methylated forms in plasma, and is degraded and metabolized to conjugated forms of caffeic acid, ferulic acid and m-coumaric acid. Life Sci 2004, 75:165-78.


